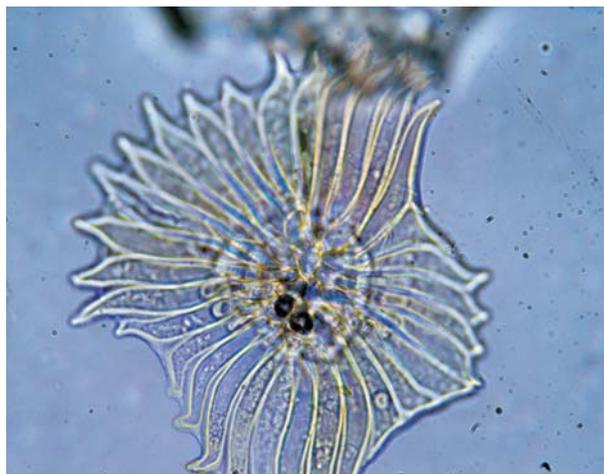
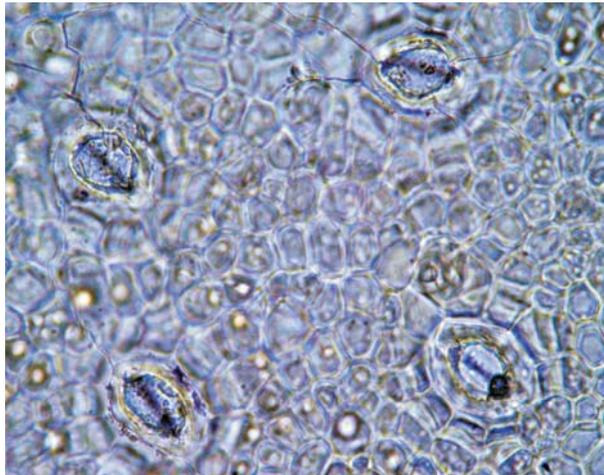


Guía ilustrada para la identificación de tejidos vegetales de plantas nativas del Chaco árido de Córdoba Argentina



Maria E. Periago & Maria Ana Dacar
2010

ÍNDICE

Introducción	2
Agradecimientos	4
Descripción técnica microhistológica	5
<i>Schinus fasciculata</i>	6
<i>Aspidosperma quebracho blanco</i>	7
<i>Bromelia urbaniana</i>	8
<i>Capparis atamisquea</i>	9
<i>Maytenus boaria</i>	10
<i>Celtis ehrenbergiana</i>	11
<i>Acacia aroma</i>	12
<i>Acacia furcatispina</i>	13
<i>Cercidium praecox</i>	14
<i>Geoffroea decorticans</i>	15
<i>Prosopis flexuosa</i>	16
<i>Prosopis sericantha</i>	17
<i>Prosopis torquata</i>	18
<i>Senna aphylla</i>	19
<i>Tricomaria usillo</i>	20
<i>Condalia microphylla</i>	21
<i>Zizyphus mistol</i>	22
<i>Castela coccinea</i>	23
<i>Lycium sp.</i>	24
<i>Lycium sp.</i>	25
<i>Aloysia gratissima</i>	26
<i>Larrea divaricata</i>	27
Bibliografía	28

Introducción

Los mamíferos herbívoros influyen sustancialmente en el funcionamiento de muchos ecosistemas debido a su impacto sobre la producción primaria, la descomposición de materia orgánica, y la redistribución de nutrientes (Batzli 1978). El flujo de materia orgánica entre organismos productores, cadenas tróficas de herbívoros y descomponedores permite el intercambio de energía y nutrientes. Según Batzli (1978), la energía y los nutrientes son factores necesarios para la producción de nueva materia orgánica a cualquier nivel trófico, y la disponibilidad de uno influye en la utilización del otro. De esta manera, los procesos interactivos en los ecosistemas son compartidos y sirven para retroalimentar el flujo de energía y reciclar los nutrientes.

Por lo tanto, los herbívoros pueden directamente influenciar la tasa de producción de su propio alimento, afectando el "fitness" de las plantas al reducir el crecimiento y la reproducción e incrementando la mortalidad (Mooney 1972). Los mamíferos herbívoros pueden cambiar la dinámica de las comunidades de plantas, alterando la diversidad de especies, la estructura tridimensional y los procesos de sucesión. Estos efectos, a su vez, afectan la diversidad y actividad de otros herbívoros, predadores y parásitos de la comunidad (Lindroth 1989).

Las hojas de una planta están compuestas por tres tipos de tejidos: la epidermis, el mesófilo y el tejido vascular o venas. La hoja se encuentra delimitada por una epidermis superior y otra inferior. Esta capa incluye diversos tipos celulares, entre los que se incluyen tricomas y estomas, con sus respectivas células guardianas (Jensen-Salisbury 1988). La técnica microhistológica utiliza estos tipos celulares para

identificar las especies vegetales consumidas por vertebrados. Las características detectadas son: presencia, tamaño y forma de tricomas y estomas; forma y organización de las células guardianas; presencia y posición de células especializadas y orientación de las nervaduras (Bauer et al. 2005).

La técnica microhistológica fue descrita por Baumgartner & Martin (1939) y desde entonces ha sido utilizada para identificar ítems de la dieta de animales (Vavra & Holechek 1980). Denham (1965) y Sparks & Malechek (1968) verificaron la técnica triturando a mano mezclas de pastos y hierbas. Desde entonces, la preparación de las muestras para identificación a través de la microscopía ha variado considerablemente. Cracker (1959) diluyó muestras fecales en agua y las colocó entre dos portaobjetos. Storr (1961) hirvió, secó y molió las muestras en una mezcla de ácidos. Después las muestras fueron lavadas con agua, teñidas de violeta, centrifugadas y montadas en portaobjetos. Otras técnicas para preparar las muestras han sido presentadas por Dusi (1949), Hercus (1960), Hegg (1961), Steward (1967), Zyznar & Urness (1969), Casebeer & Kess (1970), Ward (1970), Hansen et al. (1971) y Korfhage (1974). Aquí describimos la técnica empleada por Dacar & Giannoni (2001), que facilita la identificación de hojas y partes duras de las plantas, como ser el tallo y las semillas.

Agradecimientos

El propósito de esta guía es contribuir al análisis de la dieta de herbívoros con la técnica microhistológica. Esperamos que sea útil y ayude a ahorrar tiempo y recursos. Agradecemos a las instituciones que nos apoyaron financieramente y/o con la necesaria infraestructura: Instituto Argentino de Investigaciones de las Zonas Áridas (IADIZA), Instituto Multidisciplinario de Biología Vegetal (IMBIV), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Universidad Nacional de Córdoba, y Rufford Small Grants Foundation (RSG# 34.12.06).



*Para la versión digital y actualizaciones de la guía por favor ingresar en:
<http://personal.mendoza-conicet.gob.ar/meperiago/>

**Las fotos están sacadas en 40x, excepto las que tienen un asterisco que están en 20x. Los tallos refieren a la corteza externa de cada planta.

Técnica Microhistológica

Según Dacar y Giannoni (2001)



Para macerar los tejidos, poner la muestra en un frasco con agua y una cucharada de 17,5% NaHCO_3 (bicarbonato de sodio) y dejar reposar hasta que se ablanden. Hojas: aprox. 48 hs. Tallos y semillas: aprox. 1 semana.



Extraer la muestra y triturarla en un mortero hasta que se forme una pasta.



Añadir unas gotas de sodio hipoclorito (lavandina) para aclarar la muestra.



Volcar la muestra sobre un tamiz de 74 micrones y enjuagar con abundante agua.



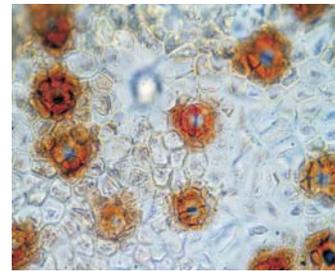
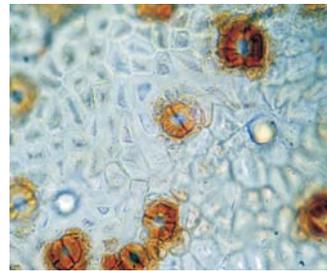
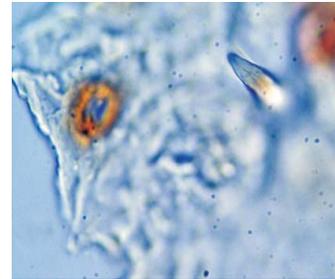
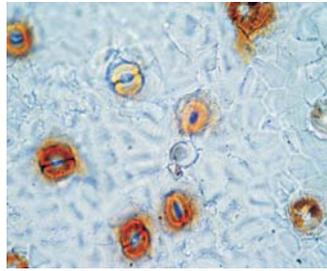
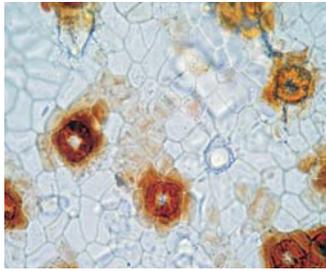
Con la ayuda de una pinza, retirar una pequeña porción de la muestra y montarla en un portaobjetos sobre una o dos gotas de glicerina. Tapar con un cubre objetos y analizar.

Familia Anacardiaceae

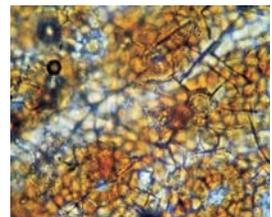
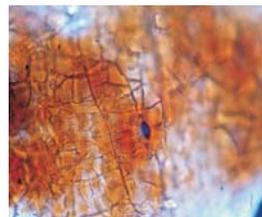
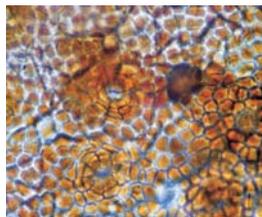
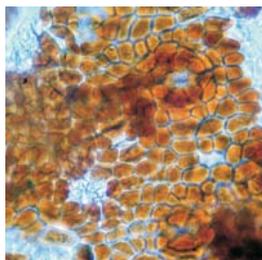
Schinus fasciculatis



Hoja

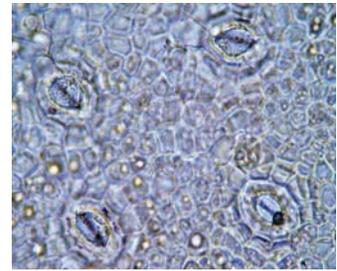
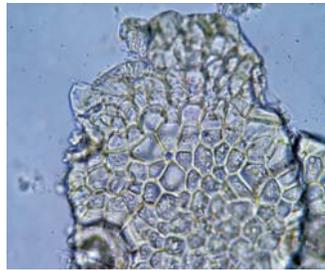
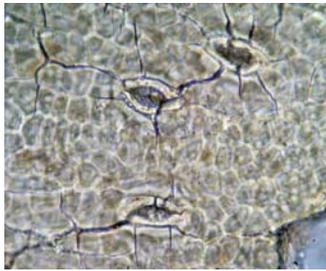


Tallo

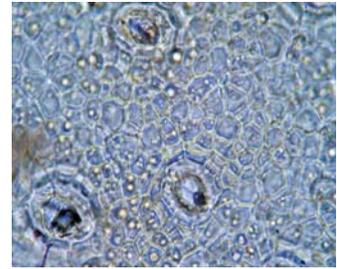
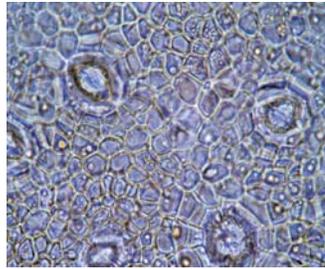
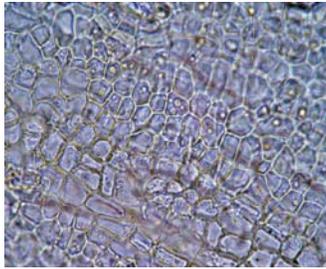


Familia Apocynaceae

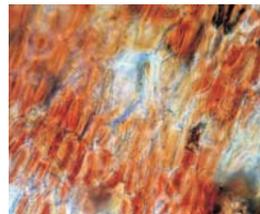
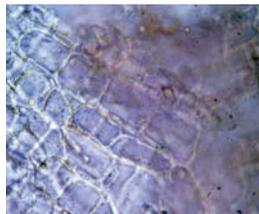
Aspidosperma quebracho-blanco



Hoja



Tallo



Familia Bromeliaceae

Bromelia urbaniana

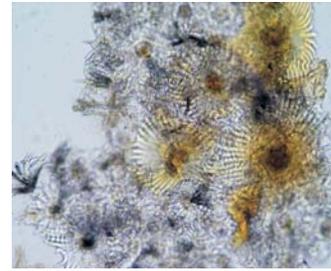
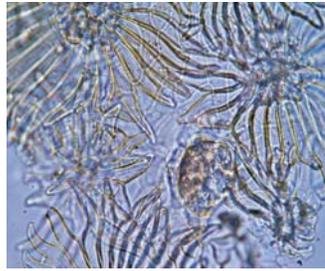
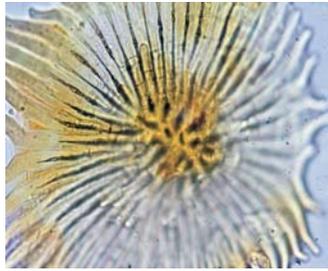


Hoja

A collection of seven microscopic images showing the internal structure of Bromelia urbaniana leaves. The images are arranged in a grid-like pattern. The top row contains two images showing vascular bundles with distinct xylem and phloem regions. The middle row contains three images showing the arrangement of vascular bundles and surrounding mesophyll cells. The bottom row contains two images showing the cellular structure of the leaf, including the epidermis and underlying tissues. The word "Hoja" is written vertically on the left side of the grid.

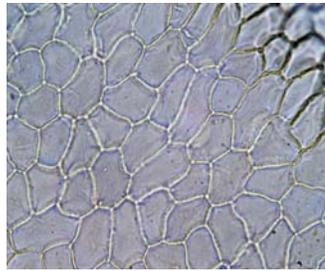
Familia Capparaceae

Capparis atamisquea

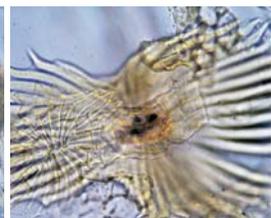
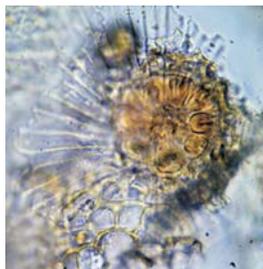


*20x

Hoja

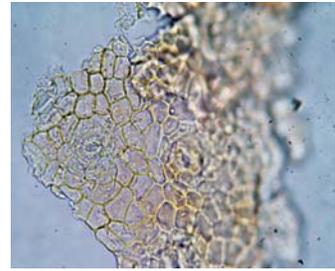
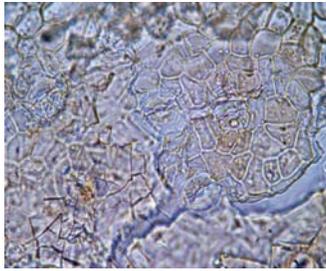


Tallo

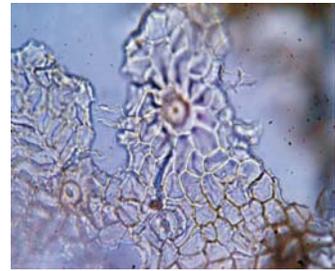
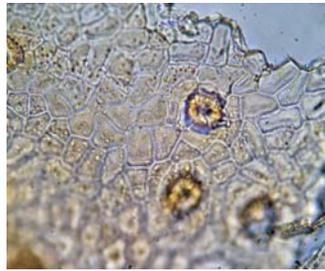


Familia Celastraceae

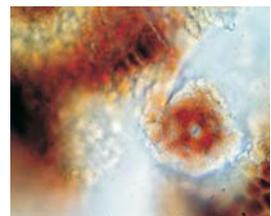
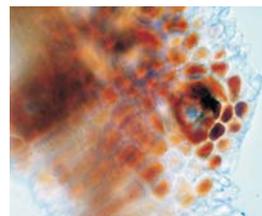
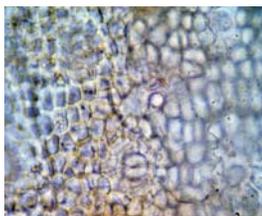
Maytenus boaria



Hoja

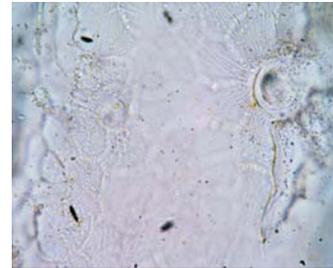
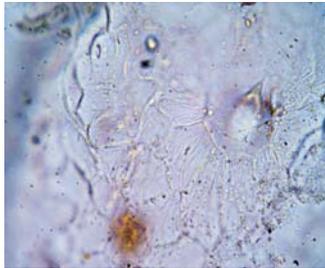


Tallo



Familia Celtidaceae

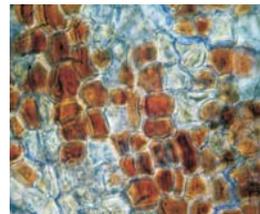
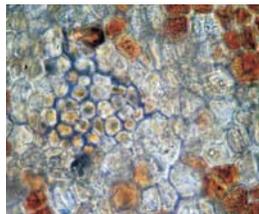
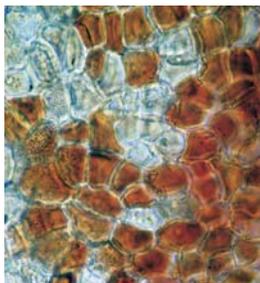
Celtis ehrenbergiana



Hoja

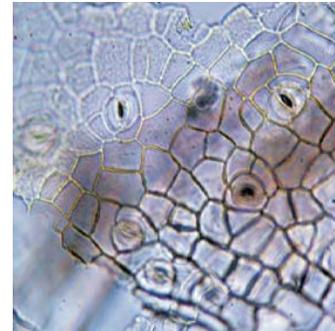
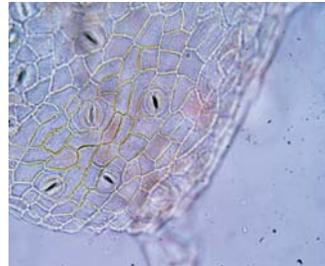
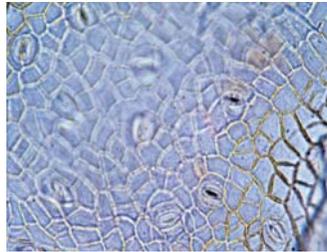


Tallo

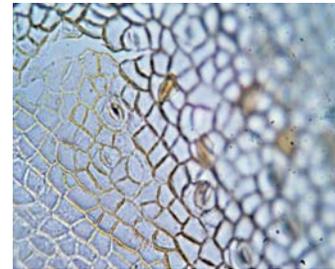
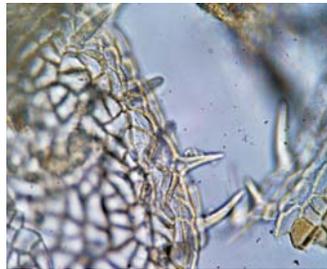


Familia Fabaceae

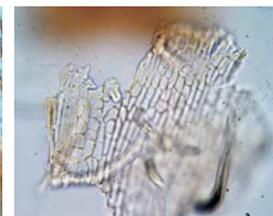
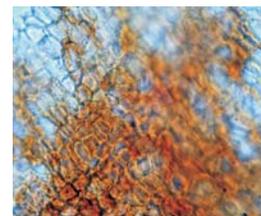
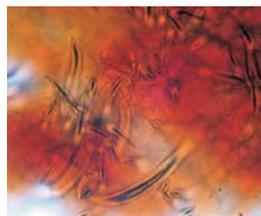
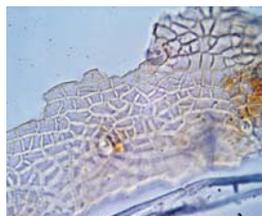
Acacia aroma



Hoja



Tallo

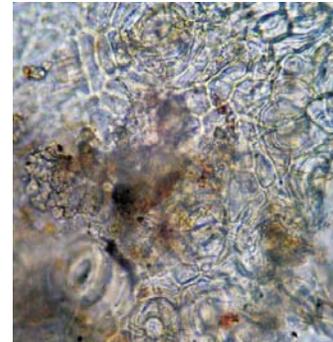
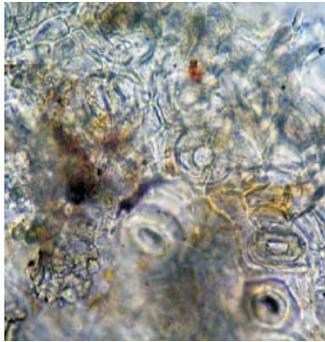


Familia Fabaceae

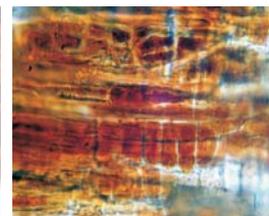
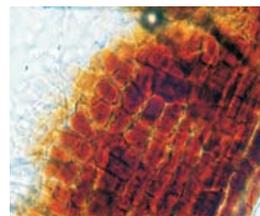
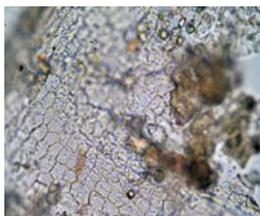
Acacia furcatispina



Hoja

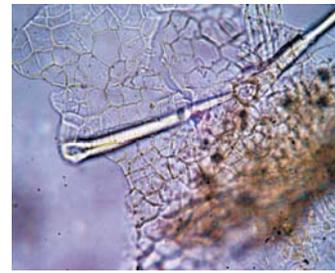
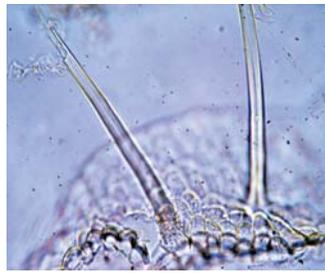
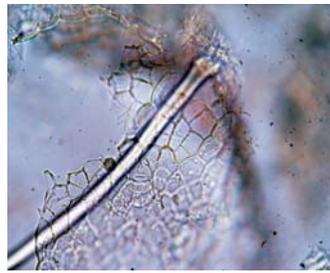


Tallo

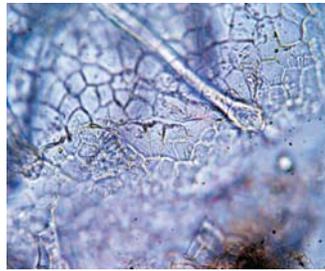


Familia Fabaceae

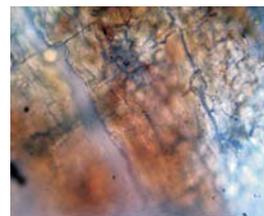
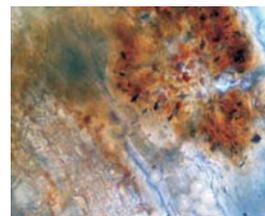
Cercidium praecox



Hoja



Tallo

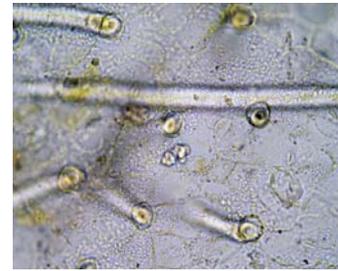
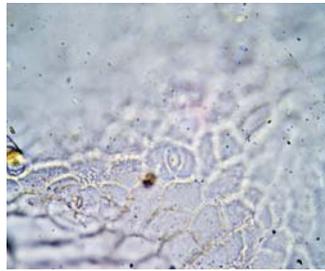
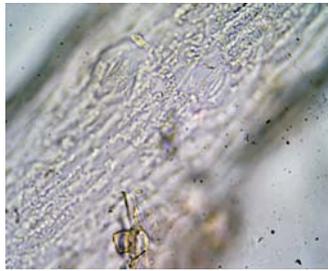


Familia Fabaceae

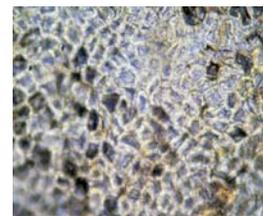
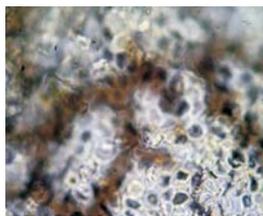
Geoffroea decorticans



Hoja

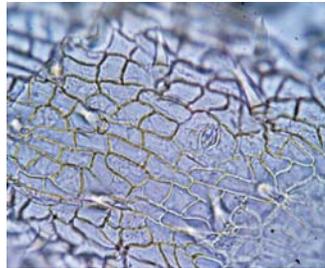
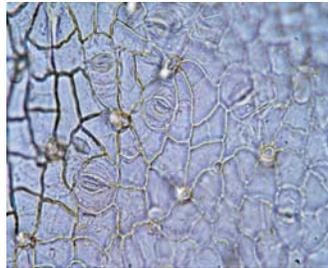


Tallo

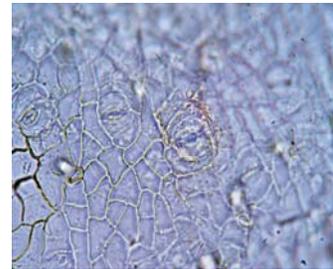
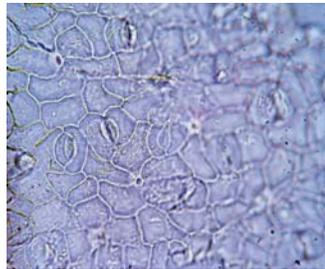
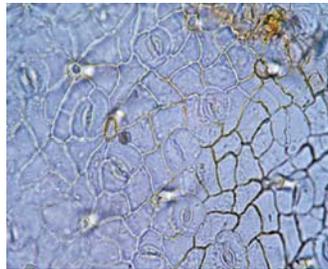


Familia Fabaceae

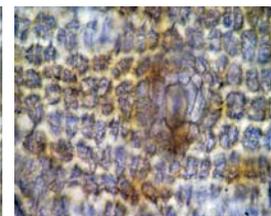
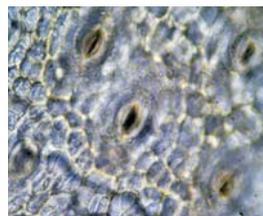
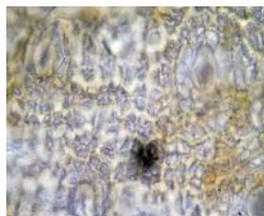
Prosopis flexuosa



Hoja

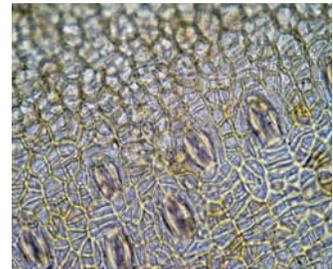
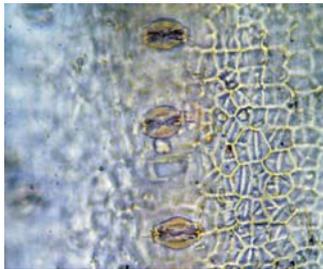


Tallo

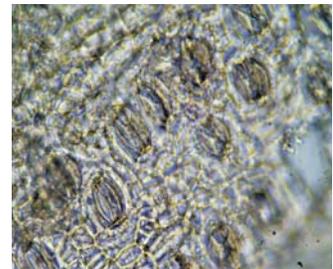
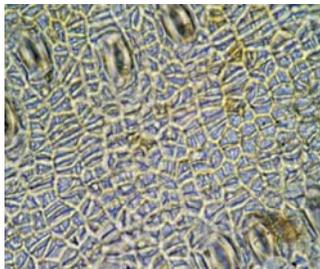


Familia Fabaceae

Prosopis sericantha

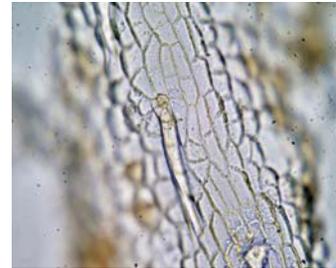
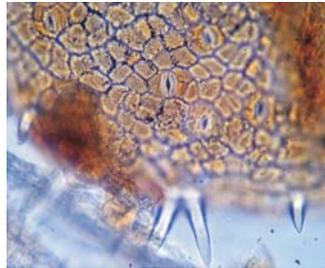
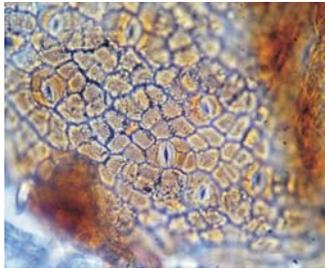


Tallo

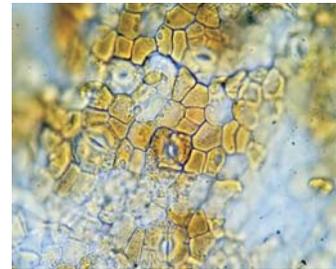
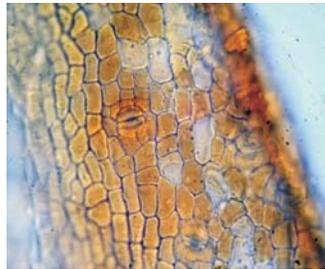


Familia Fabaceae

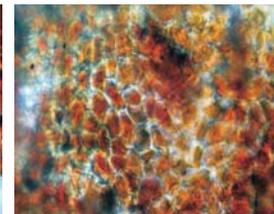
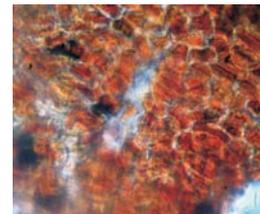
Prosopis torquata



Hoja

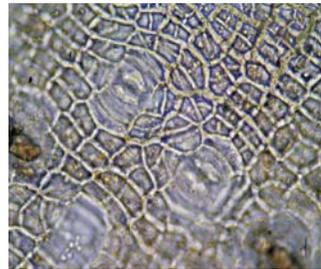
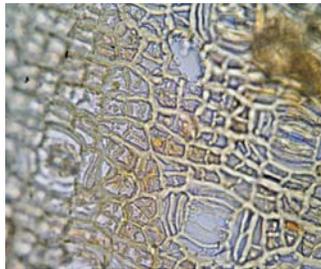


Tallo

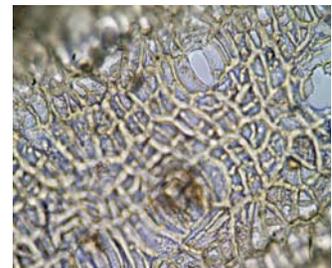
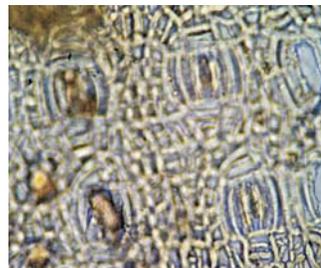
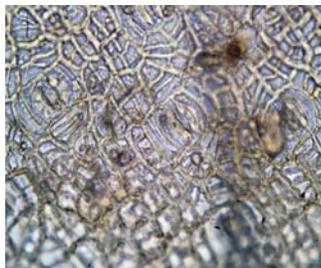


Familia Fabaceae

Senna aphylla

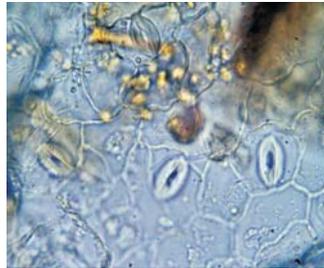
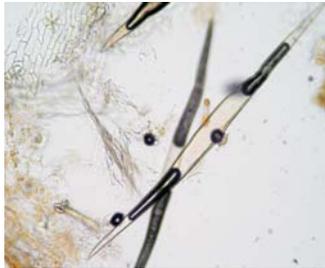


Tallo



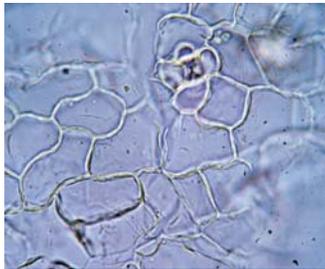
Familia Malpighiaceae

Tricomaria usillo

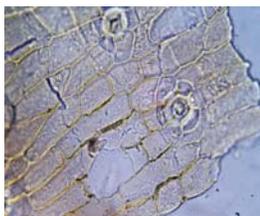


*20x

Hoja

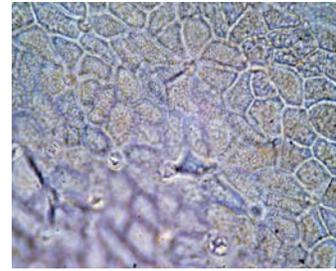


Tallo

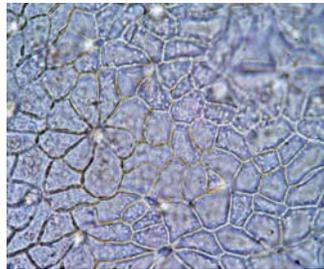
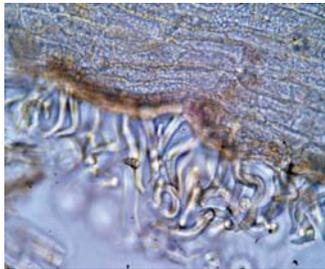


Familia Rhamnaceae

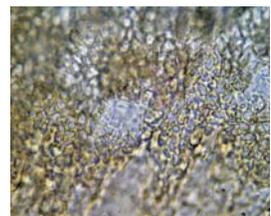
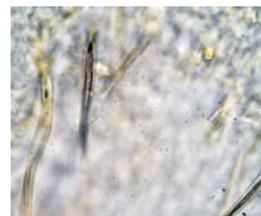
Condalia microphylla



Hoja

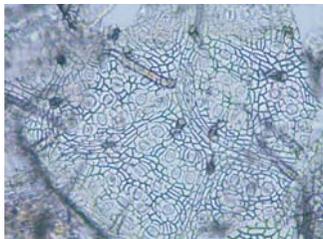


Tallo

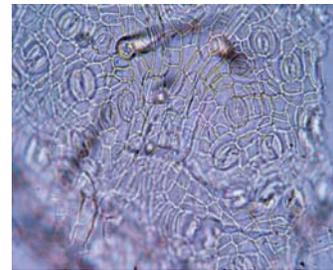
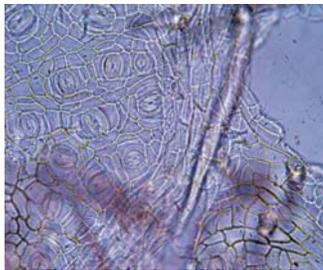


Familia Rhamnaceae

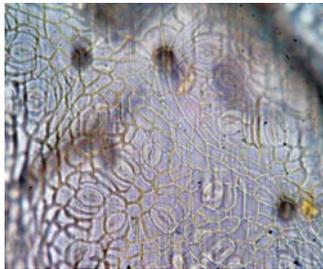
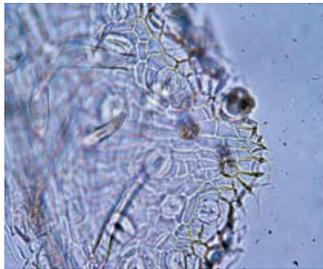
Zizyphus mistol



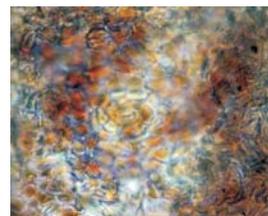
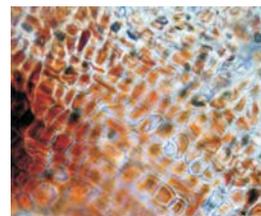
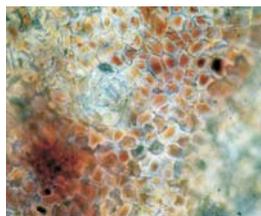
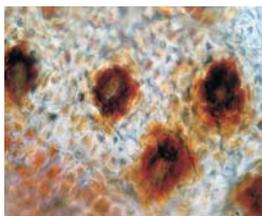
*20x



Hoja

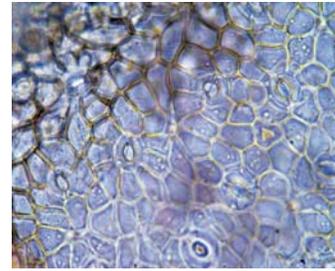
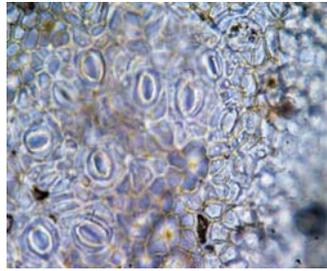
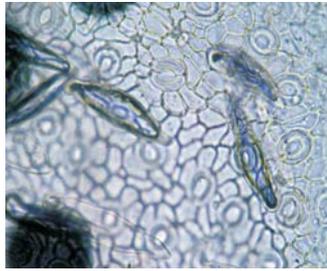


Tallo

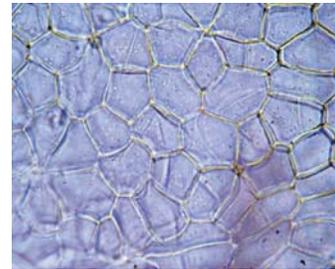
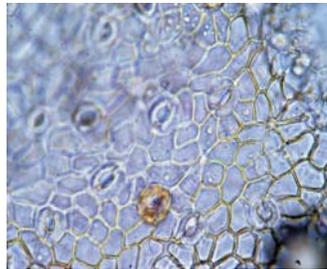
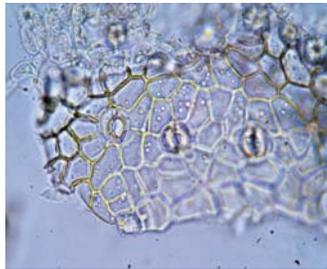


Familia Simaroubaceae

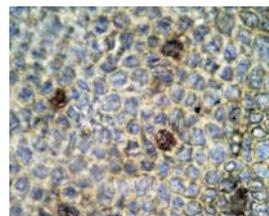
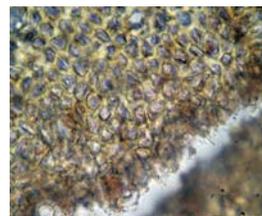
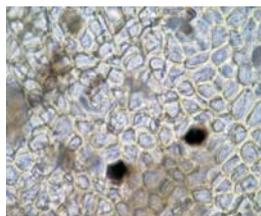
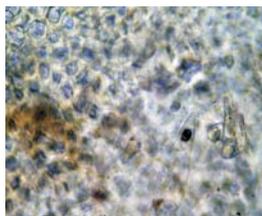
Castela coccinea



Hoja

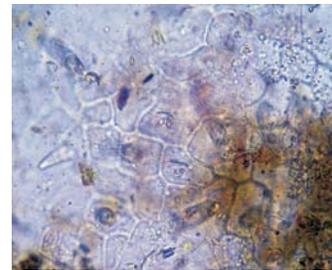
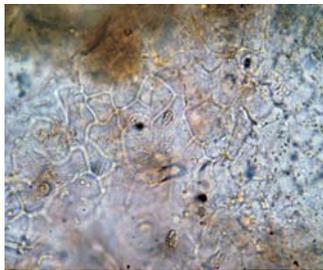


Tallo

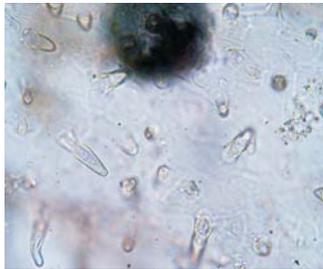


Familia Solanaceae

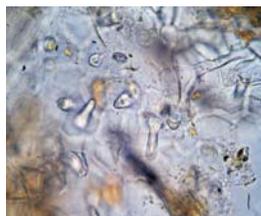
Lycium sp.



Hoja

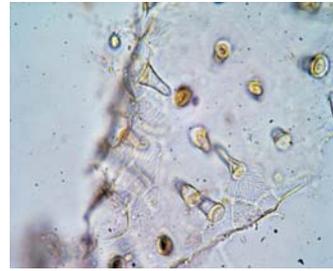
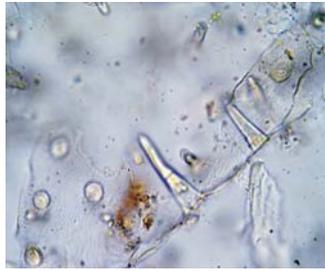
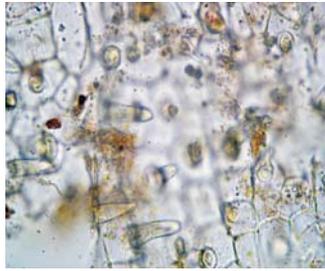


Tallo

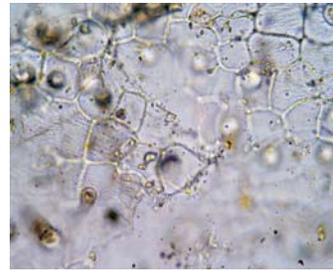
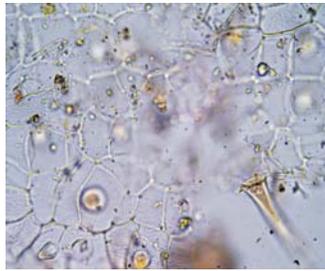
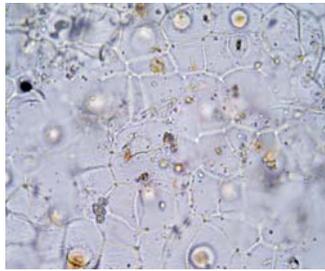


Familia Solanaceae

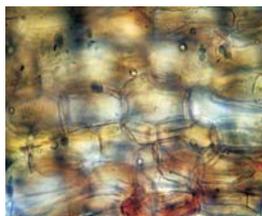
Lycium sp.



Hoja

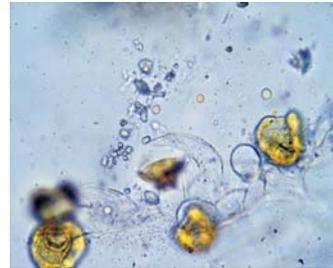
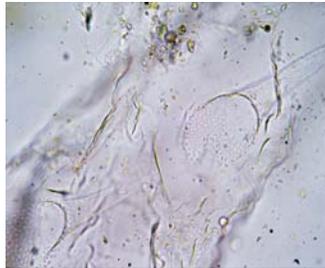
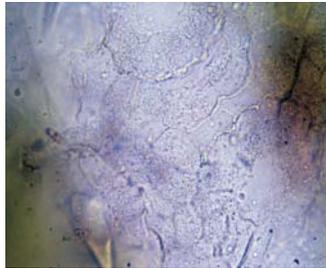


Tallo

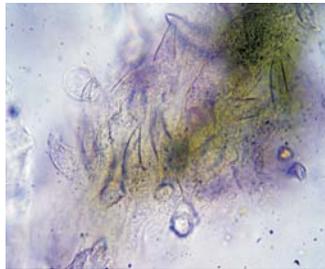


Familia Verbenaceae

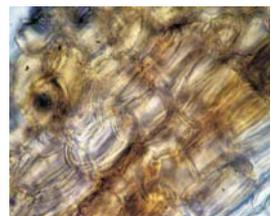
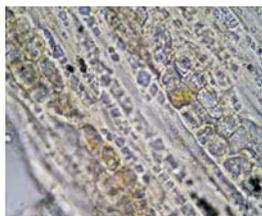
Aloysia gratissima



Hoja

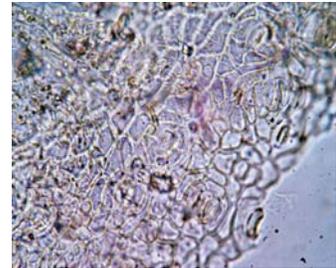
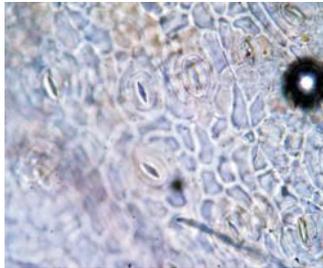
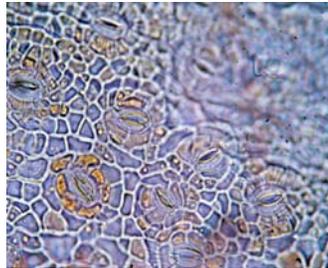


Tallo

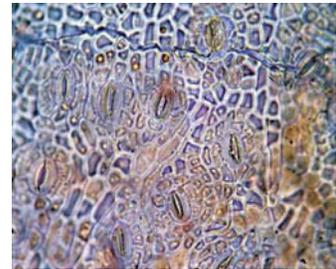
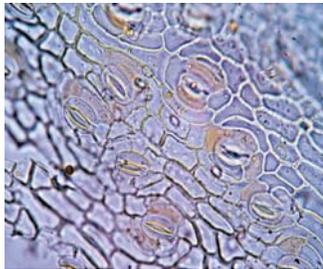
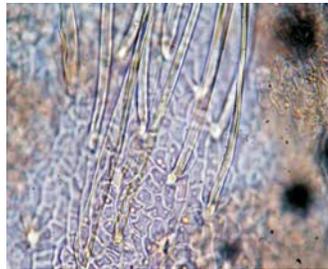


Familia Zygophyllaceae

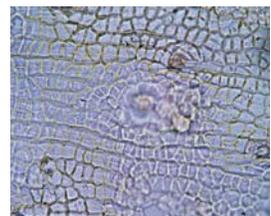
Larrea divaricata



Hoja



Tallo



Bibliografía

- Batzli, G.O. 1978. The role of herbivores in mineral cycling. En: Adriano, D.C. & I.L. Brisbin, eds. Environmental chemistry and cycling processes. ERDA Symp.Ser., U.S. Dept. of Energy, Washington D.C., p. 95-112.
- Bauer, M. de O., J.A. Gomide, E.A. Monteiro da Silva, A.J. Regazzi & J.F. Chichorro. 2005. Análise Comparativa de Fragmentos Identificáveis de Forrageiras, pela Técnica Micro-Histológica. R. Bras. Zootec., 34(6):1841-1850.
- Baumgartner, L.L., & A.C. Martin. 1939. Plant histology as an aid in squirrel food-habits studies. J. Wildl. Manage., 3: 266-268.
- Cracker, B.H. 1959. A method of estimating the botanical composition of the diet of sheep. N.Z.J. Agr. Res., 2: 72-85.
- Casebeer, R.L., and G.G. Kess. 1970. Food habits of wildebeest, zebra, hartebeest, and cattle in Kenya Masailand. E. African Wildl. J., 8: 25-36.
- Dacar, M.A & S.M. Giannoni. 2001. Technical note: A simple method for preparing reference slides of seed. J. Range Manage., 54: 191-193.
- Denham, A.H. 1965. In vitro fermentation studies on native sandhill range forage as related to cattle preference. M.S. Thesis, Colo. State Univ., Fort Collins. 78 p.
- Dusi, J.L. 1949. Methods for the determination of food habits by plant micro-techniques and histology and their application to cottontail rabbit food habits. J. Wildl. Manage., 13: 295-298.
- Hansen, R.M., A.S. Moir, and S.R. Woodmansee. 1971. Drawings of tissues of plants found in herbivore diets and in the litter of grasslands. U.S. Int. Biol. Prog. Tech. Rep. 70. 36 p.
- Hegg, O. 1961. Analysis of big game droppings to determine their dietary composition in the Swiss National Park. Revue Suisse de Zoologie, 68: 156-165.
- Hercus, B.H. 1960. Plant cuticle as an aid to determining the diet of grazing animals. Proc. 8th Int. Grassld. Cong., 443 p.
- Jensen, W.A. & F.B. Salisbury. 1988. Botánica. Libros McGraw-Hill de México, México. 762 p.
- Korfhage, R.C. 1974. Summer food habits of elk in the Blue Mountains of northeastern Oregon based on fecal analysis. MS. Thesis. Washington State University, Pullman. 117 p.
- Lindroth, R.L. 1989. Mammalian Herbivore-Plant Interactions. En: Plant-Animal Interactions. Abrahamson, W.G., ed. McGraw-Hill Inc. New York, 481 p.
- Mooney, H.A. 1972. The carbon balance of plants. Ann. Rev. Ecol. Sys. 3: 315-346.
- Sparks, D.R., and J.C. Malechek. 1968. Estimating percentage dry weight in diets using a microscopic technique. J. Range Manage., 21: 264-265.
- Steward, D.R. 1967. Analysis of plant epidermis in feces: A technique for studying the food preferences of grazing herbivores. J. Applied Ecol., 4.
- Storr, G.M. 1961. Microscopic analysis of feces, a technique for ascertaining the diet of herbivorous mammals. Aust. J. Biol. Sci., 14: 157-164.
- Vavra, M., R.W. Rice and R.M. Hansen. 1978. A comparison of esophageal fistula and fecal material to determine steer diets. J. Range Manage., 31: 11-13.
- Ward, A.L. 1970. Stomach content and fecal analysis: methods of forage identification. En: Range and Wildlife Habitat Evaluation. U.S. Forest Serv. Misc. Pub. No. 1147. 146 p.
- Zyznar, E. and P.J. Urness. 1969. Qualitative identification of forage remnants in deer feces. J. Wild. Manage., 33: 506-510.